## BEHAVIOR OF NEMATHORIN NEMATODES IN SOIL PLANTS WITH PEACH TREES AND TOMATO PLANTS INFECTED WITH NEMATODES

[8]

Weshahy, K. M.<sup>(1)</sup>; Mahgoob, A. E.<sup>(1)</sup>; Nasr, I. N.<sup>(2)</sup> and El-Sahregy, Fatma, A.<sup>(2)</sup>

1) Faculty of Agriculture, Ain Shams University 2) Agriculture Research Center

#### ABSTRACT

genera plant-parasitic nematodes, i.e. Criconemella Seven sp., Ditylenchus spp., Helicotylenchus spp., Meloidogyne spp., Pratylenchus spp, Rotylenchulus spp. and Tylenchorhynchus were recorded in the rihzospher in tomato plants. Such genera differed greatly in their prevalence and levels of abundance. The genera of Helicotylenchus spp., Pratylenchus sp. and, Meloidogyne spp. were dominant. The genera Tylenchorhynchus sp., Rotylenchulus spp., Criconemella sp. and Ditylenchus spp. were the second most important genera. On the other hand, the genera of Meloidogyne spp. and Tylenchorhynchus sp. were the most frequently occurred. Regarding Peach orchards, there were nine genera plant-parasitic nematode, i.e. Criconemella Ditylenchus Helicotylenchus spp., sp., spp., Hemicriconemoides sp. Heterodera spp., Meloidogyne spp., Pratylenchus Rotylenchulus spp. and Tylenchorhynchus spp. The genera spp., Helicotylenchus spp. and Hemicriconemoides sp. were dominant. Genera, however Pratylenchus spp .and Meloidogyne spp. were the most frequently occurred. The environmental behavior of the nematicide residues, fothiazate in sandy loam soils in peach orchards and tomatoe plants were investigated under field conditions. In relation to the degradation pattern in the selected soils, the obtained results indicated that the nematicide residues were dissipated more rapidly in the soil planted with tomatoes compared with peach orchard soils. The calculated half-life periods (t1/2) were 3.7 and 7.4 days for fosthiazate in soil planted with tomatoes and trees of peach.

Vol. 35, No.2, Sept. 2016

**Key words:** Behavior- Nemathorin-Nematodes-soil-Peach Trees-Tomato Plants.

#### **INTRODUCTION**

Nematodes are widespread and problematic on tomatoes and peach crops. Producing areas depending on soil type where they can proliferate and cause serious damages. Species of nematodes reported to be associated with tomatoes or peach crops include Rotylenchulus reniformis, Radopholus similis, Hemicriconemoides mangiferae and Meloidogyne spp. (M. incognita and *M. javanica*). General decline in tree vigor is observed in response to high nematode populations. Among surveyed soil types, sandy loam soil appeared to have population of greater numbers of root-knot nematodes, Meloidogyne spp. than clay soil. While, Helicotylenchus spp. and Rotylenchulu spp. had high density in the clay soils (Townshend, 1972; Prot and Van Gundy, 1981; Windham and Barker, 1985; Jatala, P. 1989; Ismail, 1992; McSorley, 1992; Mohamed, 1998; Tsiropoulos, et al. 2005; Korra, et al. 2009 and Osborn et al. 2009). Fosthiazate is an organophosphate compound that is currently under development as a nonfumigant nematicide. Adsorption and degradation behavior of a pesticide in soil has a strong effect on its environmental fate as well as efficacy for pest control (Qin et al., 2004). Pantelelis, et al. 2006 investigated the degradation and adsorption of the organophosphorus nematicide, fosthiazate in nine soils with various physicochemical and biological characteristics. Studies revealed the Fosthiazate concentration was strongly correlated with soil organic matter. (Kapouzas, et al. 2007) applied fosthiazate is an organophosphorus

144

Vol. 35, No.2, Sept. 2016

Weshahy et al.

nematicide which was recently included that it should be used with special care in soils vulnerable to leaching. Thus, the leaching of fosthiazate investigated in. columns packed with three different soil (Wu *et al.* 2004) determined the resestive dynamics of fosthiazate in tomato and soil was studied in this paper utilizing liquid chromatography with tandem mass spectrometry (LC-MS/MS).

#### MATERIAL AND METHODS

#### Nematode survey and extraction:

To determine the infestation level with plant parasitic nematodes infecting peach trees and tomato plants in the present study, survey of plant parasitic nematodes associated with tomato plants or peach orchards (age of tree peach about10 years) in Elhakmia village, Meet gamr destricte at El-Dakahlia governorate was carried out during 2010. Forty six samples from tomato plants and twenty two from peach trees were randomly collected from the rizhosphere using a stainless steel half-tube. The samples were kept in poly-ethylene bags and put in ice box, then transferred directly to the laboratory for nematode extraction. About 300-400 ml. of water was added to each of the soil sample (250 gm) in plastic pan for 15 min. The mixture was agitated by fingers. After a few seconds, the suspension was sieved through 200 and 400 mesh sieves (Goodey, 1957). Nematodes present in the suspension were obtained from the 400 mesh and extracted for 72 hours by the modified Baerman pans technique (Goodey, 1963). The final volume of extracted suspension was adjusted to about 50 ml. The nematodes were counted using (Hawksely) counting slide and light microscope. Nematodes

Vol. 35, No.2, Sept. 2016

were identified to genetic level according to Mai & Lyon, 1975. Population density per 250 gm of soil and Percentage of Frequency Occurrence genera in relation to surveyed soil type sites was calculated according to (Norton ,1978) as follows:-

Population density (P.D) = Total number of individuals of genus

Number of samples containing this genus

Frequency Occurrence% (F.O %) = Number of samples containing a genus X 100

Number of collected samples

## **Pesticide Used:**

The Nematicides, i.e., Nemathorin (S-sec –butyl 0-ethyl 2 –oxo-1, 3-thiazolidin-3-ylhosphonothioat) (fosthiazate) as organophosphate compound were used in this study in form of commercially granules (10% G) formulation.

## Soil type:-

Sandy loam soil was used in this investigation as soil type. The mechanical and physical and chemical characteristic of the studied soil are presented in Tables (1-2). Soil samples were period of June, analyzed according to (Dewis, J. and Freitas, F. 1970).

Soil	Measured parameters (%)						
Туре	CaCO <sub>3</sub>	<b>Coarse sand</b>	Fine Sand	Clay	Silt	Salt	
Sandy Loam	3	4.6	42.13	35	15	0.27	

Vol. 35, No.2, Sept. 2016

Weshahy et al.

Soil Type	Soluble Captions' meq/L				Soluble Anions meq/L				Soil pH
	$Mg^{++}$	С	K	$Na^+$	<b>SO</b> <sub>4</sub> <sup>-2</sup>	Cl	HCO <sub>3</sub> -	CO3 <sup></sup>	
Sandy Loam	0.43	.94	0.03	2.8	1.37	0.39	0.17		7.8

Table (2): Values of Cationic and anionic content of used soils

#### **Pesticides application:**

The pesticide application was conducted in separate areas each composed of 1600 m<sup>2</sup> of the mentioned peach and tomato locations during the period of June; 2010 each area divided into four plots each of 400 m<sup>2</sup>. For each one treatment it was distributed in a complete randomized design with three replicates. The pesticide applications was carried out by prod casting of Fosthiazate (10%G) at rate 30 kg/Fedden, Three plots were left without treatment as control. The treated soil was ploughed after pesticides application and irrigated after application by regular irrigation system (each 15 days) during the season.

#### Soil Sampling technique:

After application, forty five samples were taken from the upper 0-10 cm layer of each plot using shovel. The samples were taken after an hour, 1, 3, 9, 16, 25, 33, 48, 63 and 78 days from pesticide application. Each sample was about two kilograms which was mixed thoroughly and three representative sub-samples (each of 100 g) were taken and then transferred to the laboratory. The collected samples were left to air dryness and passed through sieves of meshes to maintain a uniform particle size and stored in clean plastic bags at  $(-20^{\circ}C)$  until residue analysis.

Vol. 35, No.2, Sept. 2016

#### **Extraction of fosthiazate**

Method described by Nelsen and Cook (1979).

**Clean-up:** The clean-up of fosthiazate from soil extracts, the dry extract was then subjected to the clean up procedure suggested by Mills *et al* (1972).

#### **Quantitaive determination:**

All the extracted and cleaned up samples were kept at  $-20^{\circ}$ C until the determination. Quantitative analysis of fosthiazate (10% G) was performed by the gas chromatograph (GC), Hp 6890 serial equipped with flame photometric detector (FPD) operated in the phosphorus mode (529 nm filter) according to the method described by Al-Samariee *et al.* (1988). The extracted samples were dissolved in ethyl acetate and injected under the following conditions: Capillary column PAS-1701 (30 m x 0.32 mm I'd x 0.25 mm film thickness). Detector temp was 250°C, injector temp was 245°C, and the column temperature was programmed 190°C and holds 2min., and rises to 260°C, at a rate of 10°C / min., and holds 8 minutes. Nitrogen carrier gas flow was 4 ml min<sup>-1</sup>., hydrogen flow was 75ml/min and air flow was 100ml/min.

Under those conditions, the retention times for, fosthiazate were 9.333 min., respectively. A series concentration of, 2.5, 5 and 10 ug per ml of ethyl acetate were prepared from fosthiazate as working standard solutions. The amounts of tested pesticide in the ample were derived from the standard calibration curves concentrated by plotting the peak areas against different concentrations of the active ingredient of each pesticide.

Vol. 35, No.2, Sept. 2016

#### **RESULTS AND DISCUSSION**

#### Frequency and population density of recovered nematode genera:

Data in Table (3) revealed the presence of 7 genera plant-parasitic nematode *i.e.* Criconemella sp., Ditylenchus spp. Helicotylenchus spp., Meloidogyne spp., Pratylenchus sp., Rotylenchulu sp and Tylenchorhynchus sp. infecting tomato plants. Such genera differed greatly in their prevalence and levels of abundance. The genera *Helicotylenchus* spp., *Pratylenchus* sp. and, *Meloidogyne* spp. were the most dominant where their population densities were 547,494 and 462 nematodes per 250 gm soil, respectively. The genera Tylenchorhynchus spp., Rotylenchulus reniformis, Criconemella sp. and Ditylenchus spp. were the second most important genera, where their population densities were 378, 280, 216 and 200 respectively. In the other hand, the genera *Meloidogyne* spp. and *Tylenchorhynchus* sp. were the most frequently occurred where their values of percentages of frequency of occurrence were 33.3 and 28.3, respectively. Regarding Peach orchards, there were 9 plant-parasitic nematode genera, *i.e. Criconemella* sp., *Ditylenchus* spp., Helicotylenchus sp., Hemicriconemoides sp. Heterodera spp., Meloidogyne spp., Pratylenchus penetrans, Rotylenchulu sreniformis and **Tylenchorhynchus** The *Helicotylenchus* sp. genera spp. and Hemicriconemoides sp. were the most dominant where their value of population density was 400 nematodes per 250 gm soil for each. While, the genera *Pratylenchus* and *Meloidogyne* spp. were the most frequently occurred where the values of percentage of frequency of occurrence were 27.3 and 13.6, respectively. These results are in agreement with the findings of Windham and Barker (1985), Ismail (1992) and Korra, et al. (2009).

Vol. 35, No.2, Sept. 2016

**Table (3):** Percentage Frequency of occurrence and Population Density in250gm of plant parasitic nematodes associated with Peach treesand tomato plants.

Host plant	Genera of Nematodes	(%)Frequency of occurrence	Population Density	
	Criconemella sp.	4.5	180	
	Ditylenchus spp.	4.5	160	
	Helicotylenchus spp.	9.1	400	
Deach	Hemicriconemoides sp.	9.1	400	
Prunusparsiag	Heterodera spp.	4.5	250	
Frunuspersica	Meloidogyne spp.	13.6	265	
	Pratylenchus spp.	27.3	190	
	Rotylenchulus spp.	9.1	200	
	Tylenchorhynchus sp.	9.1	250	
	Helicotylenchus spp.	8.7	547	
	Meloidogyne spp.	33.3	462	
Tomato	Pratylenchus spp.	10.9	494	
Lycopersicone	Criconemella sp.	7.9	216	
sculentum	Ditylenchus spp.	14.3	200	
	Tylenchorhynchus spp.	28.3	378	
	Rotylenchulus spp.	14.3	280	

Determination of the amounts of Nemathorin residue in (sandy loam soil):

Data in Table (4) and illustrated in Figures (1, 2) represent the detected residues of Nemathorin in sandy loam soil, from both the soil plants of tomatoes and Peach trees. The initial residues of nemathorin in sandy loam soil from soil Peach trees was 373.05 ppm. After the three days of application decreased rapidly to 206.9 ppm showing 44.53% loss. The decline in residual amounts were continued after 9, 16, 25, 33, 45, 63 and 78days to reach 184.58, 110.41, 98.16, 93.81, 82.93, 56.68, and 44.53 ppm with 50.52, 70.40,

Vol. 35, No.2, Sept. 2016

Weshahy et al.

73.69, 74.85, 77.77, 84.81 and 88.06 % loss, respectively. Also, the obtained results represent the residues of Nemathorin in sandy loam soil from soil plant of tomatoes. Indicated that the initial amount residue residues of Nemathorin after 3 days were 98.74 ppm



Fig. (1): Residues of (fosthiazate) in soil planted with tomatoes



Fig. (2): Residues of(Fosthiazate) in soil planted with peach

Vol. 35, No.2, Sept. 2016

Time after application (days)	soil plai	nted with to	mato	soil planted with Peach			
	Residues (mg kg <sup>-1</sup> )±SD	Loss (%)	Persistence (%)	Residues (mg kg <sup>-1</sup> )±SD	Loss(%)	Persistence (%)	
Z	237.40± 6.07	00.00	100.00	373.05±79.87	00.00	100.00	
1	117.25±5.70	50.61	49.39	229.05±27.59	38.60	61.40	
3	98.74±4.29	58.41	41.59	206.9±12.28	44.53	55.47	
9	76.46± 4.50	67.80	32.20	184.58±8.37	50.52	49.48	
16	67.74±4.53	71.47	28.53	110.41±7.68	70.40	29.60	
25	12.53± 1.4	94.72	5.28	98.16±7.63	73.69	26.31	
33	4.47± 2.85	98.12	1.88	93.81±2.78	74.85	25.15	
48	2.58±1.68	98.91	1.09	82.93±2.70	77.77	22.23	
63	2.22± 1.44	99.06	0.93	56.68±2.17	84.81	15.19	
78	1.82±0.47	99.23	0.77	44.53±1.57	88.06	11.94	
$\overline{X}$	61.355 b			132.166 a			
HL (day)	3.7			7.4			
K	.1856			.0939			

**Table (4):** Fosthiazate residues in treated sandy loam soil cultivated with tomatoes and trees of peach after several time of application

 $\overline{X}$ : The mean residues of soil. K: degradation rate.

L.S.D<sub>.05</sub> between soil planted with tomato and soil planted with peach= 11.26 days showing 58.41% loss, while after 9, 16, 25, 33, 48, 63 and 78days such residues reached 76.46, 67.74, 12.53, 4.47, 2.58, 2.22, and 1.82, respectively.

#### REFERENCES

- Al-Samariee, A. I.; Shaker, K. A. M. and Al- Bassny, M. A. (1988): Residue levels of three organophosphorus insecticides in sweet pepper grown in commercial green houses. Peptic. Sci. 22: 189-194.
- Chabrier, C; J. Hubervic and P.Quénéhervé (2002): Evaluation of fosthiazate (Nemathorin 10G) for the control of nematodes in banana fields in Martinique. Nematropica; 32(2), 137-147.

Vol. 35, No.2, Sept. 2016

- Dewis, J. and Freits, F. (1970): Physical and chemical methods of soil and water analysis.F A O-soil bulletion No-10,pp275.
- Goodey, J. B. (1963): Laboratory methods of work with plant and soil nematodes. Tech. Bull., No. 2, H. M. S. O., pp. 72.
- Goodey, T. B. (1957): Laboratory methods for work with plant and soil nematodes. Tech. Bull. No. 2, Min. Agric., Fisheries & Food, London, pp. 47.
- Ismail, S.A.A. (1992): Nematodes associated with some fruit orchards in newly reclaimed areas of Sharkia governorate. M. Sc. Thesis Fac. Agric. Zagazig Univ.
- Jatala, P. (1989): Biological Control of Plant-Parasitic Nematodes. Annual Review of Phytopathology, 24: 453-489
- Karpouzas, D.G.; Goliae, P. M. S. and Tsiropoulos, N. G. (2007): Leaching of the organophosphorus nematicide fosthiazate. Chemosphere, 68(7): 1359-64.
- Korra, A. K. M.; Gehan, A. M. Abd- El-Malek; Gomah, A. A.; Deabes, A.A. A and El-Sherif, A.A.H. (2009): Management of wilt, root rot diseases as well as root - knot nematodes in guava in Egypt. J. Biol. Chem. Environ. Sci., 4(4): 443-471.
- Mai, W. F. and Lyon, H. H. (1975): Pectoral key to genera of plant- parasitic nematodes. Cornell Univ. Press. Thecae, New York nematodes. Wiley, New York, pp. 268.
- Mills, P. A.; Bong, B. A.; Kamps, L. R. and Burke, J. A. (1972): Elution solvent system for florisil column clean up in organochlorine pesticide residue analysis J.A.O.A.C,55, (1): 39-43.
- McSorley, R. (1992): Nematological problems in tropical and subtropical fruit tree crops. Nematropica, 22, No (1):103-116.
- Mohamed, E. A. and Mohamed. J. (1998): Comparative assessment of pasteuria penetrate pentanes and three nematicides for the control of Meloidogyne javanica and their effect on yields of successive crops of tomato and melon. Fundam. Appl Nemalol: 21,(2),113-118.

Vol. 35, No.2, Sept. 2016

- Nelsen, T. R. and R. F. Cook (1979): A high-pressure liquid chromatographic method for the determination of carbofuran residues in soil and water. J. Agric. Food Chem. 27 (6): 1186-1188.
- Norton, D.C. (1978) Ecology of plant parasitic nematodes. Wiley, N.Y., 268.
- Osborn, R. K.; S.G. Edwards; Wilcox, A. and Haydock, P. PJ. (2009): Potential enhancement of degradation of the nematicides aldicarb, oxamyl and fosthiazate in UK agricultural soils through repeated applications. Pest Management Science, 66 (3): 229–344.
- Pantelelis, I.; Karpouzas, D. G.; U. Menkissoglu-Spiroudi and Tsiropoulos, N. (2006): influence of soil physicochemical and biological properties on the degradation and adsorption of the nematicide fosthiazate. J. Agric. Food Chem.: 54 (18): 6783–6789.
- Prot. J. C. and Van Gundy, S. D. (1981): Effect of soil texture and the clay component on migration of M. incognita second stage juveniles. J. Nematol., 13: 213-217.
- Qin, S.; Gan, J.; W. Liu and Becker, J.O. (2004): Degradation and adsorption of fosthiazate in soil. J .Agric Food.chem. 52(20): 6239-42.
- Tsiropoulos, N.G.; Likas, D. T. and Karpouzas, D. G. (2005): Liquid chromatographic determination of fosthiazate residues in environmental samples and application of the method to a fosthiazate field dissipation study. J AOAC Int.;88(6):1827-33.
- Townshend, J. L. (1972): Influence of edaphic factors on penetration of corm roots by Pratylenchus pentrans and P. minyus in three Outa of soil. Nematologica, 18: 201-212.
- Windham, G. L. and Barker, K. R. (1985): Reproductive and damage potential of Meloidogyne incognita host races on soybeab. J. Nematolo., 17: 516.
- Woods, S. R.; Haydock, P. P. J. and Edmunds, C. (1999): Mode of action of fosthiazate used for the control of the potato cyst nematode (Globodera pallida,)Annals of Applied Biology,135,(1): 409–415.
- Wu, J.; Wang, K. and Zhang, H. (2012): Residues and Dissipation Dynamics of Fosthiazate in Tomato and Soil) Bull Environ Contam Toxicol

Vol. 35, No.2, Sept. 2016

89:664–668.

Wu, M and Hu, J. (2014): Residue analysis of fosthiazate in cucumber and soil by QuEChERS and GC-MS.Chemical Papers:68,(10): 1368-1374.

# سلوك المبيد النيماتودى نيماثورين في تربة مزروعة بأشجار الحوج ونباتات الطماطم المصابة بالنيماتودا

[^]

قدرى محمود وشاحي<sup>(۱)</sup>- أحمد عيد عبد المجيد محجوب<sup>(۱)</sup>- إسلام نعمان نصر<sup>(۲)</sup> فاطمة عبد الله فكري السهريجي<sup>(۲)</sup> ۱) كلية الزراعة، جامعة عين شمس ۲) مركز البحوث الزراعية

## المستخلص

تم حصر أهم اجناس النيماتودا النباتيه الموجوده في تربه طفليه رمليه، بمركز ميت غمر محافظة الدقهليه مزروعه بنباتات الطماطم (قبل المعاملة) ووجد بها ۷ أجناس هي:

Criconemella, Ditylenchus, Helicotylenchus, Meloidogyne, Pratylenchus penetrans, Rotylenchulus reniformis and Tylenchorhynchus .

وهذه الاجناس اختلفت فى النسبه المئويه للتعاقب ومستوى الظهور . وكانت الأجناس الأكثر سياده وظهوراً في التربة هي:

Helicotylenchus spp., Pratylenchus sp. and, Meloidogyne.

يليها أجناس: Rotylenchulus, Tylenchorhynchus وفي حالة تربه أشجار الخوخ كانت أهم الأجناس الأكثر سيادة وظهوراً في التربة هي: Hemicriconemoides, Helicotylenchus.

بينما أجناس: Meloidogyne, Pratylenchus هي الأكثر سيادة وظهوراً وتعاقباً. ترجيب

تم إجراء رصد لمتبقيات هذا المبيد وإجراء التحليل الإحصائي والرياضي له بعد التطبيق. أنثاء الفتره من شهر يونيه وحتى شهر أغسطس من عام ٢٠١٠. وقد تم إجراء الرصد بعد مرور ساعة زمنية واحدة ٣، ١٩، ٢، ٢٥ ، ٢٣ ، ٤٨ ، ٣٣، ٧٨ يوم من تطبيق المبيد. وقد أشارت النتائج المتحصل عليها بأنه يوجد فرق معنوي مقداره ١١،٢٦ يوم عند تطبيق المبيد المشار اليه، بالاضافه الي أنه كان أكثر اختفاءاً بالتربه المزروعه بالطماطم، عن التربه المزروعه بالخوخ، وذلك يرجع الى زيادة عدد مرات الرى المستخدمه لنباتات الطماطم، عن عد مرات الرى المستخدمه لرى اشجار الخوخ.

Vol. 35, No.2, Sept. 2016